

UNIVERSITY OF ESWATINI
FACULTY OF SCIENCE AND ENGINEERING
DEPARTMENT OF BIOLOGICAL SCIENCES
MAIN EXAMINATION PAPER 2020/2021

COURSE CODE: BIO451
TITLE OF PAPER: BIOINFORMATICS
TIME ALLOWED: **THREE (3) HOURS**
INSTRUCTIONS: NUMBERS IN BRACKETS DENOTE THE NUMBER OF MARKS
THIS PAPER COMPRISES OF **TWO SECTIONS** and **ONE ANNEX**.
SECTION A COMPRISES OF EIGHT (8) QUESTIONS.
SECTION B COMPRISES OF SEVEN (7) QUESTIONS.
ANSWER ALL QUESTIONS IN SECTIONS A AND B.

TOTAL MARKS AVAILABLE: 70

NO ADDITIONAL MATERIAL (E.G. NOTES, BOOKS, PHONES, SMART WATCHES ETC) MAY BE TAKEN INTO THE EXAMINATION.

THE USE OF A SCIENTIFIC CALCULATOR IS PERMITTED.

DO NOT OPEN THIS PAPER UNTIL PERMISSION HAS BEEN GRANTED BY THE CHIEF INVIGILATOR

Section A

Total marks available: 35

A poacher has been arrested by the police in possession of several processed animal parts. Unfortunately, it is not possible to identify the animal(s) just by looking at them. The police have sent samples of the processed animal parts to you for forensic analysis. You have extracted and sequenced the samples using your next-generation sequencer.

AQ1: State what **type of molecule** and **file format** of the data you expect to receive from the sequencer. [2]

AQ2 Describe what you would do with the sequence data to help identify the organism(s). In your answer, include what databases you would use. [4]

AQ3: The results have come back for one of the samples. Explain what the four columns inside the black oval (Query cover, E value, Per. Ident and Accession) tells you about your search.

Descriptions		Graphic Summary	Alignments	Taxonomy								
Sequences producing significant alignments					Download	▼	▼	▼	Show	100	▼	▼
<input type="checkbox"/> select all 0 sequences selected												
Description	Scientific Name	Common Name	Taxid	Max Score	Query Score	Query Cover	E value	Per. Ident	Acc. Len	Accession		
<input type="checkbox"/> Tragelaphus strepsiceros isolate PHC11 mitochondrion, complete genome	Tragelap... greater...		9946	3195	3195	100%	0.0	99.77%	16382	JH632708.1		
<input type="checkbox"/> Tragelaphus spekei mitochondrion, complete genome	Tragelap... Sitatunga		69298	2809	2809	100%	0.0	95.75%	16408	EF636357.1		
<input type="checkbox"/> Tragelaphus oryx isolate PHC13 mitochondrion, complete genome	Tragelap... eland		9945	2776	2776	100%	0.0	95.42%	16397	JH632704.1		
<input type="checkbox"/> Taurotragus derbianus mitochondrion, complete genome	Taurotra... giant el...		303930	2776	2776	100%	0.0	95.46%	16399	EF536354.1		
<input type="checkbox"/> Tragelaphus eurycerus isolate SUW8 mitochondrion, complete genome	Tragelap... bongo		69297	2771	2771	100%	0.0	95.36%	16349	JH632703.1		
<input type="checkbox"/> Tragelaphus scriptus isolate MBP11 mitochondrion, complete genome	Tragelap... bushbuck		66440	2748	2748	100%	0.0	95.18%	16384	JH632706.1		
<input type="checkbox"/> Tragelaphus scriptus isolate PhC17 mitochondrion, complete genome	Tragelap... bushbuck		66440	2732	2732	100%	0.0	95.01%	16402	JH632707.1		
<input type="checkbox"/> Tragelaphus buxtoni mitochondrion, complete genome	Tragelap... mountai...		69296	2723	2723	100%	0.0	94.80%	16601	HC_038064.1		
<input type="checkbox"/> Tragelaphus scriptus isolate CAR31 mitochondrion, complete genome	Tragelap... bushbuck		66440	2553	2553	99%	0.0	93.26%	16401	JH632705.1		
<input type="checkbox"/> Tragelaphus angasi isolate MBP15 mitochondrion, complete genome	Tragelap... nyala		66437	2514	2514	100%	0.0	92.77%	16399	JH632702.1		
<input type="checkbox"/> Tragelaphus imberbis mitochondrial complete genome	Tragelap... lesser k...		9947	2507	2507	100%	0.0	92.62%	16401	LH873135.1		
<input type="checkbox"/> Tragelaphus imberbis mitochondrion, complete genome	Tragelap... lesser k...		9947	2507	2507	100%	0.0	92.62%	16405	EF536356.1		
<input type="checkbox"/> Tragelaphus imberbis mitochondrial 12S and 16S ribosomal RNAs and Phe-, Val- and Leu-tRNA genes	Tragelap... lesser k...		9947	2507	2507	100%	0.0	92.62%	2686	LH6493.1		
<input type="checkbox"/> Boselaphus tragocamelus mitochondrion, complete genome	Boselap... nilgai		9947	2318	2318	100%	0.0	90.73%	16322	EF536350.1		

[2]

AQ4: Based on the results above, what accession number would you pursue? [1]

AQ5: After further investigation, using GenBank, you come across this result (see Annex 1) for a second sample. Identify the following:

- a) The size and type of molecule [2]
- b) The full name of the GenBank Division [2]
- c) The protein name [1]
- d) Organism (common and scientific) [3]
- e) Name of the gene encoding for this protein [2]

- f) The accession number of the gene encoding for this protein [2]

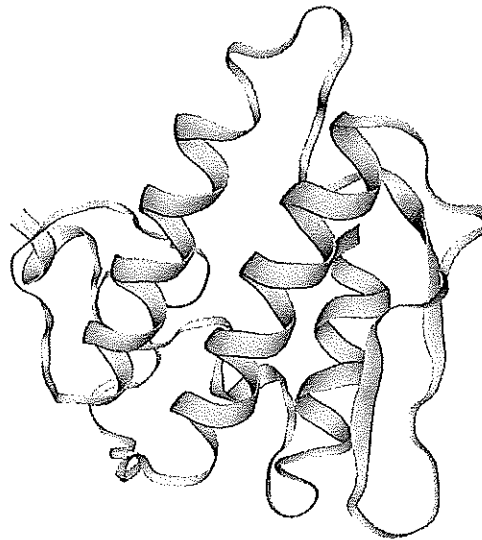
AQ6: You have sent your results to the legal teams handling the case. Unfortunately, there is some dispute over the results from the second sample (AQ5). Both the prosecution and defense teams have requested some further analysis. Specifically, they request you to analyse the sample that they gave you against the very same organism identified in from Annex 1. Explain how you could go about getting a tissue sample from the organism (in Annex 1) to run your investigation.

[5]

AQ6: Describe how you would use bioinformatics to determine the structure and function of the protein (from AQ5), including the database(s) you would use. [5]

AQ7: List two reasons for determining the structure of proteins. [2]

AQ8: Describe the key feature(s) of the protein below. [2]



Section B

Total marks available: 35

BQ1: Which BLAST application would you use to identify the following sequences:

- a) AGAAGTTCCAGATCATTCCAAGA [1]
- b) AUCCCAGGGUUCACCAGAGCCAGGAGA [1]
- c) KAGTAKCATVADDCHUABCCAGAAGNMWSS [1]
- d) VLPWGQMSFWGATVITNLLSAIPYIGNTLV [1]

BQ2: Identify which of the following sequence(s) would you enter into the tBLASTn database: [1]

- a) MGGNTAKCRRADDCHUABCCAGAAGNMWSS
- b) CGAGAUUAUCCCAGGGUUCACCAGAGA
- c) TAGGTTCCAGATCATTCCAAGA
- d) ATPYISSAFVGYVLPWGPTVQMSFWGATVITNLLS

BQ3: What is the default scoring matrix for BLASTp? [1]

BQ4: Assuming a scoring method: match = +1, mismatch = -2, and gap = -1, using the Needleman-Wunsch algorithm:

a) Initialize the scoring matrix below.

[2]

		T	C	T	G	A	T	T	C	T	C
T											
C											
G											
C											
A											
T											
C											
A											
C											
C											

b) Calculate the score of the matrix and fill the traceback matrix. Use arrows to indicate how you determined the path forward. Use circles and arrows to determine your trace back. [5]

c) Deduce the best alignment based on the traceback matrix. [3]

BQ5: Assuming a scoring method: match = +1, mismatch = -2, and gap = -2, using the Smith and Waterman algorithm:

a) Initialize the scoring matrix below.

[2]

		T	C	T	G	A	T	T	C	T	C	C
T												
A												
G												
C												
A												
T												
T												
A												
G												
C												

b) Calculate the score of the matrix and fill the traceback matrix. Use arrows to indicate how you determined the path forward. Use circles and arrows to determine your trace back. [5]

c) Deduce the best alignment based on the traceback matrix. [3]

d) Were there any sub-optimal alignments? Explain your answer. [2]

BQ6: Align the following two sequences using a dot plot

[3]

- TCTGATTCTC
- TCGATCAC

BQ7: You have four sets of nucleic acid sequences of very different lengths to align. Which type of algorithm(s) would be most appropriate? Explain your answer and give ONE advantage for your algorithm(s).

[4]