| TITLES OF PAPER: | SEPARATION METHODS \& ENYIRONMENTAL <br> ANALYTICAL TECHNIQUES |
| :--- | :--- |
| COURSE CODES : | C611 \& ERM642 |
| TIME ALLOWED : | THREE(3) HOURS |
| INSTRUCTIONS : | ANSWER ANY FOUR(4) <br> QUESTIONS. EACH QUESTION <br>  <br> $\quad$CARRIES 25 MARKS. |

> A PERIODIC TABLE AND OTHER USEFUL DATA HAVE BEEN PROVIDED WITH THIS PAPER

REQUIRED: GRAPH PAPER

DO NOT OPEN THIS QUESTION PAPER UNTL PERMISSION TO DO SO HAS BEEN GRANTED BY THE CHIEF INVIGILATOR

## Question 1 (25 marks)

(a) For the extraction of a weak acid, HB into an organic phase, the acid being monomeric and its anion being insoluble in the organic phase, employ the basic equilibria involved to:
(i) Obtain the expression for the distribution ratio, D , in terms of $\mathrm{K}_{\mathrm{a}}, \mathrm{K}_{\mathrm{DHB}}$ and $\left[\mathrm{H}_{3} \mathrm{O}^{+}\right]$.
(ii) Show how a linearized form of the expression for D above can be used in evaluating the values of $\mathrm{K}_{\mathrm{a}}$ and $\mathrm{K}_{\mathrm{DHB}}$ graphically.
(b) (i) The distribution coefficient, $\mathrm{K}_{\mathrm{D}}$, of an organic compound between water and an organic solvent is 18.0 . If 100 mL of an aqueous solution of the compound, buffered at pH 6.00 is extracted three times with 50 mL of the organic solvent, calculate the percentage remaining in the aqueous phase. ( $\mathrm{K}_{\mathrm{a}}=2.0 \times 10^{-6}$ ).
(ii) What will be the value of D at pH 4 ?
(iii) Comment on the results of (i) and (ii) above.
(e) A certain metal ion $\mathrm{M}^{\mathrm{n+}}$ is extracted by a chelating agent. The concentration of the chelating agent is 0.010 M and the following data are obtained:

| pH | 1 | 2 | 3 | 4 | 5 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| D | $10^{-8}$ | $10^{4}$ | 1 | $10^{4}$ | $10^{8}$ |

From the plot of $\log \mathrm{D}$ vs. pH obtain the values n and K (collection of constants)

## Question 2 ( 25 marks)

(a) (i). State the expression that relates the net retention volume, $\mathrm{V}_{\mathrm{n}}$, and the specific retention volume, $\mathrm{V}_{\mathrm{g}}$ and define the other parameters in it. (2)
(ii) What are the factors that influence the value of $\mathrm{V}_{g}$ in a solvent and what assumption is made in this respect?
(iii) Assuming an ideal behaviour, how is the net volume affected by an increase in the temperature and volatility of the solutes?
(b). A $5.00-\mu \mathrm{L}$ sample containing aniline $\left(\mathrm{C}_{6} \mathrm{H}_{5} \mathrm{NH}_{2}\right)$ and anisole $\left(\mathrm{C}_{6} \mathrm{H}_{5} \mathrm{OCH}_{3}\right)$ together with other substances was injected into a GC. The heights for the peaks of these two solutes in the resulting chromatogram were 4.22 (aniline) and 7.60 (anisole) chart divisions. Another $5.00-\mu \mathrm{L}$ sample was injected together with $0.25 \mu \mathrm{~L}$ of pure aniline (all in the same syringe), producing aniline and anisole peak heights of 8.73 and 7.60 chart divisions. Calculate the concentration; in volume $\%$, of the two components under the following assumptions:
(i) The detector responds equally to both compounds.
(ii) The detector response (on a volume basis) is 1.35 times more for anisole than for aniline
(C) (i) Give the two expressions for the resolution, $\mathrm{R}_{5}$ of two adjacent peaks in a chromatogram. Account for the factors that influence its value.
(ii) Using a 2.0 m column, what height of a theoretical plate is needed to achieve a resolution of 1.0 ? (Given that $\alpha=1.05$, and $\mathrm{k}^{\prime}=0.5$ ).

## Question 3 ( 25 marks)

(a) Explain the term 'band broadening' in GC analysis.
(b) With reference to the van Deemter equation, account for the contribution by each of the factors responsible for band broadening and column efficiency in terms of HETP.
©. By making reference to the Van Deemter equation, predict the effect (increase, no effect, cannot determine), on the plate height, H , in each of the following conditions, with only one parameter varied at a time:
(i) Decreasing the particle size.
(ii) Increasing the column temperature.
(iii) Increasing the thickness of the liquid coating material.
(iv) Increasing the linear gas flow rate.
(d). The analysis of $n$-hexane was carried out by injecting $2-\mu \mathrm{L}$ samples unto a 3.00 m GC column. The following table contains the data obtained:

| Sample | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Flow Rate <br> (mL/s) | 2.00 | 1.51 | 1.20 | 1.05 | 0.84 | 0.67 | 0.53 | 0.43 |
| Retention time, <br> $\mathbf{t}_{s}(\mathrm{~s})$ | 329.4 | 382.2 | 430.2 | 457.2 | 517.2 | 589.8 | 678.6 | 761.4 |
| Peak Width (s) | 21.0 | 23.4 | 25.8 | 28.2 | 32.4 | 40.8 | 48.6 | 57.0 |

(i) Prepare a van Deemter plot (i.e. plot HETP vs. flow rate).
(ii) Determine the optimum flow rate
(iii) Calculate N and H (HETP), at the optimum flow rate.

## Question 4 ( 25 marks)

(a) For the HPLC technique, distinguish between:
(i) Analytical and guard columns.
(ii) Normal phase chromatography and reverse phase chromatography.
(iii) Isocratic elution and gradient elution.
(b) Give four of the basic requirements for a HPLC pump.
(c) Describe two of the different kinds of pumps commonly employed in HPLC. What are the advantages and disadvantages of each?
(d) During the HPLC determination of the caffeine in an analgesic tablet, a $10-\mu \mathrm{L}$ injection loop was used in preparing the caffeine standards. The following data were obtained:

| [Standard], <br> (ppm) | 50.0 | 100.0 | 150.0 | 200.0 | 250.0 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Signal(arbitrary <br> units) | 8354 | 16925 | 25218 | 33584 | 42002 |

A single analgesic tablet was placed in a small beaker, dissolved with 10.0 mL of methanol and the contents, including the binder, were transferred into a $25-\mathrm{mL}$ volumetric flask and diluted to level with methanol. The sample was then filtered and a 2.50 mL aliquot was transferred into another $25-\mathrm{mL}$ volumetric flask, diluted to level with methanol and analyzed similarly as the standards. A signal of 21469 was obtained for the caffeine. Estimate the amount of caffeine (in mg), in the analgesic tablet. (6)

## Question 5 (25 marks)

(a) Define the following terms and discuss the factors that influence their values:
(i) Electrophoretic mobility.
(ii) Electroosmotic flow velocity.
(6)
(b) Give a brief account of the principles of capillary zone electrophoresis(CZE). What is its main limitation and how is it overcome by the micelar electrokinetic chromatography(MEKC)?
(C) Discuss the efficiency and solute resolution of capillary electrophoresis, indicating the parameters that influence them.
(d) CZE was employed for the analysis of $\mathrm{NO}_{3}{ }^{-}$in aquarium water, using $\mathrm{IO}_{4}{ }^{-}$as an internal standard. Standard solutions of $30.0-\mathrm{ppm} \mathrm{NO} 3_{3}{ }^{\circ}$ and 20.0 ppm of $\mathrm{IO}_{4}{ }^{-}$gave peak heights (arbitrary units), of 190.0 and 200.2 respectively. A 2.50 mL water sample from an aquarium was transferred into a 250.0 mL volumetric flask and then diluted to volume after adding sufficient internal standard to make its concentration 10.00 ppm . Analysis gave signals of 29.2 and $105.8 \mathrm{NO}_{3}{ }^{-}$and $\mathrm{IO}_{4}{ }_{4}$, respectively. Estimate the concentration of $\mathrm{NO}_{3}{ }^{-}$in the aquarium sample in ppm .

## Question 6(25 marks)

(a) What are the advantages of supercritical fluid chromatography over GC and. HPLC?
(b) Discuss the principles of size exclusion chromatography. What are the likely sources of error during the application of this technique for analysis?
© Distinguish gel - filtration from gel - permeation exclusion chromatography.
(d) During a gel - permeation experiment, a $2.5 \times 50-\mathrm{cm}$ Sephadex $\mathrm{G}-200$ column was used, giving the following elution data:

| Compound | Molecular Wt.(M.W.) | Retention volume <br> $(\mathrm{mL})$ |
| :---: | :---: | :---: |
| Sucrose | 342 | 242 |
| Glucagon | 5,500 | 233 |
| Cytochrome C | 11,000 | 214 |
| Chymotripsinogen | 24,000 | 188 |
| Bovine serum albumin | 80,000 | 149 |
| Aldolase | 153,000 | 127 |
| $\alpha-$ Conaractin | 486,000 | 92 |
| $\alpha-$ Crystallin | 825,000 | 78 |
| Blue dextran | $2,000,000$ | 75 |
| Unknown | $?$ | 119 |

(i) Plot an appropriate graph from the data.
(ii) Determine the M.W. of the unknown from your plot.
(iii) Why is the retention volume nearly independent of the M.W. at both very low and very high M.W's?
(iv) Estimate the retention volume for an enzyme with M.W. of 39,000 .
(v) Calculate N (the number of theoretical plates), for two peaks having $\mathrm{R}_{\mathrm{s}}=1$ and retention volumes of $200-\mathrm{mL}$ and $205-\mathrm{mL}$.


## PERIODIC TABLE OF ELEMENTS

GROUPS


